**Solanum lycopersicum** (tomato) ethanol extract elicits anti-inflammatory effects via the nuclear factor kappa B pathway and rescues mice from septic shock

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**Abstract:** *Solanum lycopersicum*, commonly known as tomato, is widely used in raw, cooked, or liquid forms because it contains nutritional compounds that are beneficial for human health, including carotenoids, lycopene, ascorbic acid, vitamins, and minerals. The tomato is perhaps the most widely studied fruit, especially with respect to its cardioprotective effects. In this study, we aimed to identify the anti-inflammatory mechanisms by which the tomato elicits its anti-inflammatory properties. We treated murine macrophage RAW 264.7 cells with a tomato ethanol extract and performed various biochemical assays including nitric oxide inhibition, cell viability, RNA extraction, expression of pro-inflammatory mediators and cytokines, and immunoblotting, as well we assessed cell survival rates. Our results have shown for the first time that a tomato ethanol extract treatment can suppress nitric oxide production in a dose-dependent manner without cytotoxicity. Moreover, it inhibits the expression of pro-inflammatory mediators and cytokines and elicits its anti-inflammatory effects via the nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) and mitogen-activated protein kinase (MAPK) pathways. In addition, administration of tomato syrup potently rescued mice from septic shock induced by lipopolysaccharide injection. Collectively, our results elucidate details regarding the anti-inflammatory mechanisms of tomato.

**Keywords:** anti-inflammation, cytokines, pro-inflammatory mediators, septic shock, tomato ethanol extract

**Introduction**

The growing incidence of cardiovascular, immunosuppressive and chronic inflammatory diseases due to the rapid pace of industrialization poses a serious threat for the well-being of mankind. The increased amounts of social, personal, and workplace stress cannot be ignored as factors predisposing humans to premature aging, mental illnesses and early deaths [19]. Rapid and ongoing global research towards a cure for various chronic diseases has altogether contributed positively towards the health of mankind. However, the serious disadvantages of these chemical orientated medicines cannot be ignored. In order to avoid the side effects of these chemical medications, scientists all over the world are struggling to improve health conditions by using natural products like fruits and vegetables. *Solanum lycopersicum* commonly known as tomato is widely consumed as fruit and vegetable all over the world. The anti-oxidant, anti-inflammatory, anti-diabetic, and anti-lipidemic activities of the tomato have been studied extensively in the past and currently [1, 9]. The protective properties of tomato are mainly attributed to the carotenoid, lycopene, which is present in great quantities within the fruit. There is much ongoing research on the tomato as a whole fruit, or isolated lycopene, which is the most abundant and active component of the tomato. Lycopene administration alleviated prostatitis in a manner comparable to that of ciprofloxacin [14]. Furthermore it reduced the chemical and histological symptoms in iodoacetamide induced colitis in rats [28]. Anthocyanins from tomatoes have also been shown to exhibit inhibitory effects on inducible nitric oxide synthase (iNOS) and cyclooxygenase-2 (COX-2) [12].

Inflammation is the hallmark of almost every disease and if not controlled at proper time point, can lead to chronic diseases. The production of pro-inflammatory cytokines when the body encounters a foreign invader is a primary response of immune cells [21]. These pro-inflammatory cytokines and mediators further recruit more potent chemicals and cells, such as natural killer cells, that neutralize and destroy the for-
eign particle. Macrophages are the cells that phagocytize foreign particles and signal the recruitment of other cells via production of inflammatory mediators like iNOS, COX-2 and pro-inflammatory cytokines including interleukin (IL)-1β, IL-6 and tumor necrosis factor (TNF)-α [29]. Production of pro-inflammatory mediators leads to the activation of pro-inflammatory cytokines [8]. All of these chemicals when released synchronously, fight against foreign materials and alleviate infection through the nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) and mitogen-activated protein kinase (MAPK) pathways [30].

Previously we have shown the antiplatelet effects of the tomato extract [16], but here in this study, we geared to elucidate the detailed anti-inflammatory mechanisms of whole tomato ethanol extract in vitro using RAW 264.7 cells and in vivo using a septic shock ICR mouse model. It is a well-known and alarming fact that the cause of most post-operative mortalities or bacterial infections is due to severe septic shock which is defined as the systemic distribution of bacterial toxins in body leading to death [22].

Materials and Methods

Materials
Dulbecco’s modified Eagle’s medium (DMEM) and fetal bovine serum (FBS) were purchased from WELGENE (Korea). Total RNA extraction kit was purchased from Invitrogen (USA). Oligo dT, iNOS, COX-2, TNF-α, IL-6, and IL-1β primers were obtained from Bioneer (Korea). Lipopolysaccharide (LPS; Escherichia coli 055: B5) and 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazoliumbromide (MTT) were purchased from Sigma Aldrich (USA). Specific antibodies used against phospho- and/or total form of extracellular signal-regulated kinase (ERK), c-Jun N-terminal kinase (JNK), p38, nuclear factor of kappa light polypeptide gene enhancer in B cells inhibitor (IκB), iκB kinase (IKK) α/β, NF-κB p65 and β-actin as well as rabbit HRP linked secondary antibody were purchased from Cell Signaling Technology (USA). All other reagents and chemicals were obtained from Sigma Aldrich.

Animal studies
Male ICR mice (26–29 g) were purchased from Charles River, Orient Biotechnology, Gyeonggi-do, South Korea. The mice were housed in a specific pathogen free barrier facility at 21 ± 2°C with a relative humidity of 60 ± 10% under a 12 h light and dark cycle. Feed and water were provided ad libitum. All animal care and experimental procedures were approved by Animal Care Committee (2015-0062) of Kyungpook National University, Daegu, South Korea. The mice were divided into 3 groups with each group (n = 10) for survival study. Group 1 was taken as control or vehicle treated group. Group 2 was LPS control group and Group 3 was treated with water soluble tomato concentrate (DSM Nutritional Product; DSM, Switzerland) [16], which we named as tomato syrup for 3 days prior to LPS injection orally at a dose of 900 mg/kg once a day based on the equivalent human consumption of tomato per day which is 3 g/ day. The amount of lycopene present in the tomato syrup was around 3–5 mg/100 g fresh tomatoes [6, 32]. After the pretreatment of mice with tomato syrup for three days, at the fourth day groups 2 and 3 were given LPS intraperitoneally at 30 mg/kg and then the survival rate was monitored for 96 h.

Tomato ethanol extract (TEE) preparation
Whole dried tomatoes were condensed using 70% ethanol in a 20-fold volume. After condensation, the ethanol extract was filtered through a 2 µm pore Whatman filter paper. The filtrate was then frozen at ~70°C for 2 days and then vacuum dried to obtain a powered form. The powdered form was then weighed and diluted in dimethyl sulphoxide (DMSO) and used accordingly for each experiment.

Cell culture
RAW 264.7 cells, murine macrophage cell line, originating from American Type Culture Collection (ATCC-TIB-71) were cultured in complete DMEM supplemented with 5% FBS, penicillin (100 IU/mL) and streptomycin sulfate (100 µg/mL) in humidified 5% CO2 incubator at 37°C.

Nitric oxide assay
Nitric oxide (NO) measurements were performed on the basis of Griess reaction. In short, RAW 264.7 cells were seeded in 96-well plates and incubated with or without LPS (0.1 µg/mL) in the absence or presence of TEE at concentrations of 250–1,000 µg/mL for 18 h. Next day, the culture supernatants (100 µL) were mixed with Griess reagent (0.2% naphthyl ethylene diamine dihydrochloride, and 2% sulphanilamide in 5% phosphoric acid) in double distilled water at equal volumes and incubated for 5 min at room temperature. The absorbance in each well was then read at 540 nm in enzyme-linked immunosorbent assay reader (VersaMax ELISA Microplate Reader; Molecular Devices, USA).

Cell viability assay
Cytotoxic effects of TEE were examined using MTT reagent which was added to culture medium at a final concentration of 0.1 mg/mL in 96-well plate. After 4 h of incubation at 37°C in 5% CO2, the violet coloured crystals were dissolved in DMSO (100 µL/ well) and absorbance values were read at 560 nm (VersaMax Microplate Reader; Molecular Devices).

RNA extraction and quantitative real-time polymerase chain reaction (qRT-PCR)
RAW 264.7 cells were pre-treated with or without TEE at indicated concentrations for 30 min and then stimulated with LPS (0.1 µg/mL) for 18 h in 6-well plates. Total RNA was extracted using a TRIZOL reagent (Invitrogen, USA) following the manufacturer’s instructions. Subsequent steps for
cDNA for qRT-PCR were according to previous study [31]. Quantitative PCR primer sequences are given in Table 1.

**Table 1. Primer sequences used for polymerase chain reaction**

<table>
<thead>
<tr>
<th>Gene</th>
<th>Primer</th>
<th>Oligonucleotide sequence (5’-3’)</th>
</tr>
</thead>
<tbody>
<tr>
<td>GAPDH</td>
<td>F</td>
<td>5’CAATGAATACGGCTACAGCAAC3’</td>
</tr>
<tr>
<td></td>
<td>R</td>
<td>5’AGGGAGATGCTCAGTGGTG3’</td>
</tr>
<tr>
<td>iNOS</td>
<td>F</td>
<td>5’CCCTCCGAAGTTTCTGGCAGACGC3’</td>
</tr>
<tr>
<td></td>
<td>R</td>
<td>5’GGCTGTCAGAGCTCGGTGTTTGG3’</td>
</tr>
<tr>
<td>COX-2</td>
<td>F</td>
<td>5’-TCTCAGCACCACCCCCGCTCA-3’</td>
</tr>
<tr>
<td></td>
<td>R</td>
<td>5’-GCCCCCAGACCCCTGTGCA-3’</td>
</tr>
<tr>
<td>IL-1β</td>
<td>F</td>
<td>5’CAGGGTGCTGCTGCCGTCCTTC3’</td>
</tr>
<tr>
<td></td>
<td>R</td>
<td>5’TGCTTCCAACCTTTGACCTGCG3’</td>
</tr>
<tr>
<td>TNF-α</td>
<td>F</td>
<td>5’TTGACCTCAAGCCTGATGTTG3’</td>
</tr>
<tr>
<td></td>
<td>R</td>
<td>5’CCTGAGAGACGTACGTCAG3’</td>
</tr>
<tr>
<td>IL-6</td>
<td>F</td>
<td>5’-GTACTCCAGAGACCAAGAGG-3’</td>
</tr>
<tr>
<td></td>
<td>R</td>
<td>5’-TGCTGAGACAACCGCC-3’</td>
</tr>
</tbody>
</table>

F, forward; R, reverse.

**Western blot analysis**

Cytosolic and nuclear proteins were extracted according to manufacturer’s instructions using NE-PER nuclear and cytosolic extraction reagents (No. 78833 and No. 78835; Thermo Scientific, USA) from RAW264.7 cells when they were treated or untreated with TEE (250–1,000 µg/mL) in the presence or absence of LPS (0.1 µg/mL) in 6-well plates. Protein quantification was then performed using PRO-MEASURE assay kit (iNtRON Biotechnology, Korea). Proteins were then loaded onto 10% acrylamide gels, separated by SDS-PAGE and transferred onto PVDF membranes (Immobilon-P; Millipore, USA). Nonspecific binding on PVDF membranes was minimized with a blocking buffer containing 5% non-fat dry milk and 0.1% Tween-20 in Tris-buffered saline. The membranes were then incubated with specific primary antibodies overnight at 4°C followed by 1 h incubation with HRP-conjugated anti-rabbit antibody (1:3,000 dilution, Cell Signaling Technology). Bound antibodies were visualized using enhanced chemiluminescence (SUPEX ECL solution; Neuronex, Korea) and images were analyzed using ImageJ 2 software (National Center for Biotechnology Information, USA). β-actin was used as an internal control.

**Statistical analysis**

Data are presented as mean ± SEM. One-way analysis of variance followed by Dunnett’s -test was used for statistical analysis. SAS 9.3 (SAS Institute, USA) was applied for analysis. P values less than 0.01 were considered statistically significant.
Fig. 2. TEE suppressed pro-inflammatory mediators. RAW 264.7 cells were preincubated with TEE for 30 min and then stimulated with LPS for 18 h. Total RNA was extracted and mRNA expression of inducible nitric oxide synthase (iNOS) and cyclooxygenase-2 (COX-2) was determined both by real-time (A) and reverse transcriptase polymerase chain reaction (B). GAPDH was used as an internal control. Image is representative of three independent experiments. Values in bar graphs are mean ± SEM of three independent experiments. ***p < 0.001 and **p < 0.05 are considered significant compared to LPS group only.

Fig. 3. Expression of pro-inflammatory cytokines was diminished by TEE. For mRNA expression, RAW 264.7 cells were pre-treated with TEE for 30 min and then stimulated with LPS for 18 h. Total RNA was extracted and mRNA expression of interleukin (IL)-1β, IL-6 and tumor necrosis factor (TNF)-α was determined by real-time PCR (A) and RT-PCR (B). GAPDH was used as an internal control. Image is representative of three independent experiments. Values in bar graphs are mean ± SEM of three independent experiments. ***p < 0.001 and **p < 0.05 are considered significant compared to LPS group only.
Results

**TEE inhibited LPS induced inflammation in RAW 264.7 cells**

NO is produced when bacterial lipopolysaccharides bind to the Toll like receptors. Therefore, we sought to check the inhibition of NO by TEE. As shown in Fig. 1A, TEE significantly suppressed the NO production in a dose dependent manner without any cytotoxicity (Fig. 1B).

**Suppression of pro-inflammatory mediators by TEE**

In our result, we found that TEE suppressed the expression of pro-inflammatory mediators that are iNOS and COX-2 at transcriptional level as shown in Fig. 2A and B.

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**Fig. 4.** TEE elicits anti-inflammatory effects via the nuclear factor kappa B (NF-κB) and MAPK pathways. For the protein expression, RAW 264.7 cells were treated with TEE and stimulated with LPS 30 min later. Nuclear and cytoplasmic proteins were extracted by NE-PER Nuclear and Cytoplasmic Extraction Kit (Thermo Scientific). β-actin was used as an internal control. Inhibition in the phosphorylation of all downstream NF-κB (A). Suppression in the phosphorylation of MAPK factors downstream (B and C). Image is representative of three independent experiments. Values in bar graph are mean ± SEM of three independent experiments. ***p < 0.001, **p < 0.05 and *p < 0.01 are considered significant compared to LPS group only.
particularly the lycopene content exhibited strong anti-inflammatory effects act via the NF-κB and MAPK pathways. These results clearly showed that TEE and tomato syrup increased the survival rates of mice dramatically as shown in Fig. 5.

Diminution of pro-inflammatory cytokines expression levels by TEE
As shown in Fig. 3A and B, TEE had dose dependently diminished the transcriptional expression levels of the major pro-inflammatory cytokines such as IL-1β, IL-6 and TNF-α.

Signal transduction of TEE via the NF-κB and MAPK pathways
Most extracts or compounds that exhibit anti-inflammatory effects act via the NF-κB and MAPK pathways. As shown in Fig. 4A-C, TEE in a potent manner decreased the phosphorylation of all the downstream factors of NF-κB and MAPK pathways. These results clearly showed that TEE and particularly the lycopene content exhibited strong anti-inflammatory effects through which inflammatory mediators, cytokines and downstream regulating factors were suppressed.

TEE increased the survival rate in septic shock mice
The above results were the elucidation of TEE anti-inflammatory effects in vitro. This persuaded us to examine the in vivo effects of tomato syrup supplementation in mice challenged with a lethal dose of LPS i.e. 30 mg/kg. Our results showed that the tomato syrup administered orally at dosage of 900 mg/kg increased the survival rates of mice dramatically as shown in Fig. 5.

Discussion
Inflammation is the phenomenon that is characterized by the microcirculation of blood cells, primarily the white blood cell types to counteract the foreign particle. This system works in harmony with many interleukins, cytokines and chemokines to bring the cellular state back to its normal condition [21]. Generally inflammation is characterized by the five cardinal signs: swelling, pain, redness, oedema and loss of sensation at the site. These outward clinical manifestations appear because of the internal battle between the foreign particle and the body’s immune cells. Whenever a foreign particle invades a cell, a number of inflammatory mediators and cytokines are released that modulate or neutralize the invader [10]. NO is a gas that is released when L-arginine is oxidized by nitric oxide synthase (NOS) enzyme. It then mediates the production of pro-inflammatory cytokines to effectively recruit the other immune cells for defence mechanism [18]. The production of this gas is generally beneficial as it recruits the other pro-inflammatory mediators, however if uncontrolled, it leads to the excessive stimulation of pro-inflammatory mediators like iNOS and COX-2 and cytokines, IL-1β, IL-6, and TNF-α [13, 18].

Lycopene is a naturally occurring carotenoid synthesized in plants but not in animals or humans. It is abundantly found in tomatoes and various other fruits and vegetables. The lycopene content in the tomato can range from 8.8-42 µg/g of its wet weight and nearly 85 % of lycopene for dietary proportion is obtained from the tomato and tomato products [1, 27]. Numerous epidemiological studies have shown that populations consuming more tomatoes or those that have tomato as an essential component in their diets exhibit lower incidences of chronic inflammatory diseases [2, 26]. Up till now there are many studies investigating tomato as a modulator of inflammation primarily due to the lycopene content and also because lycopene targets a wide range of molecular cell machinery [4, 20]. As a result of NO production, transcriptional factors like iNOS and COX-2 are activated signaling the release of pro-inflammatory cytokines [34]. Our results (Figs. 1-3) have shown that TEE dose dependently inhibited the production of NO as well as the expression levels of iNOS, COX-2, IL-1β, IL-6 and TNF-α. All these factors are the pro-inflammatory mediators that start the inflammatory cascade or pro-inflammatory cytokines that recruit the other inflammatory cells and activate downstream signalling pathways [23, 25, 35].

After investigating the preliminary suppression of these pro-inflammatory mediators and cytokines, we further moved to examine the pathways that modulate these effects. The first one to our knowledge was the NF-κB pathway. This is a classical pathway of inflammation and is activated when the LPS binds to the Toll-like receptor 4, leading to the disassociation, translocation and activation of its downstream components like interleukin-1 receptor-associated kinase (IRAK-1), transforming growth factor β-activated kinase (TAK-1), IKKβ/α, IκB/α and finally NF-κB in the nucleus [11, 30, 33]. Our results in (Fig. 4A) have shown that TEE inhibited the phosphorylation of all of these downstream factors in the RAW 264.7 cells indicating that lycopene component of the tomato was indeed taken into the cell demonstrating its anti-inflammatory effects under LPS stimulation [24].

MAPK pathway is a stress activated pathway. It is always activated whenever the cell experiences stress either endogenous or exogenous. [7, 17, 31] Since LPS is a bacterial
toxin, it created stress in RAW 264.7 cells, leading to the pro-inflammatory activation of this pathway. Again, the lycopene content of tomatoes as shown in (Fig. 4B-C) has dose dependently diminished the phosphorylation of all the down-stream factors associated with this pathway (i.e. MAPK kinase MEK, MKK, JNK, ERK and P38). These results indicate that TEE extract exerts its anti-inflammatory effects via the NF-κB and MAPK pathways. The biggest concern of practical medical science is the avoidance of systemic inflammation that occurs as a result of post-operative sepsis or some other chronic inflammatory disease [15]. A number of studies have indicated that tomato extract or tomato products have been beneficial in the treatment of carrageenan-induced paw oedema in rat, prostatitis and colitis in rats [3, 5, 9, 23, 28]. However, no study to this extent factors associated with this pathway (i.e.

In conclusion, TEE had shown outstanding effects as an anti-inflammatory agent in RAW 264.7 cells by suppressing the production of pro-inflammatory mediators and cytokines via the NF-κB and MAPK pathways. Moreover tomato syrup dramatically increased the survival rates of mice with LPS-induced septic shock mouse model.

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